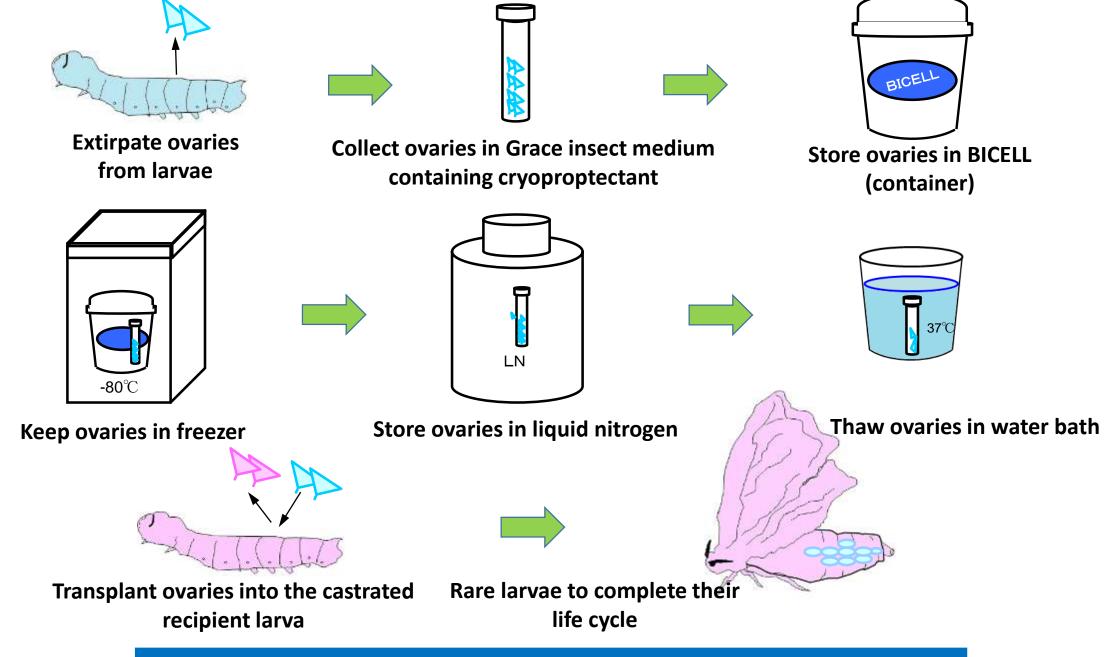
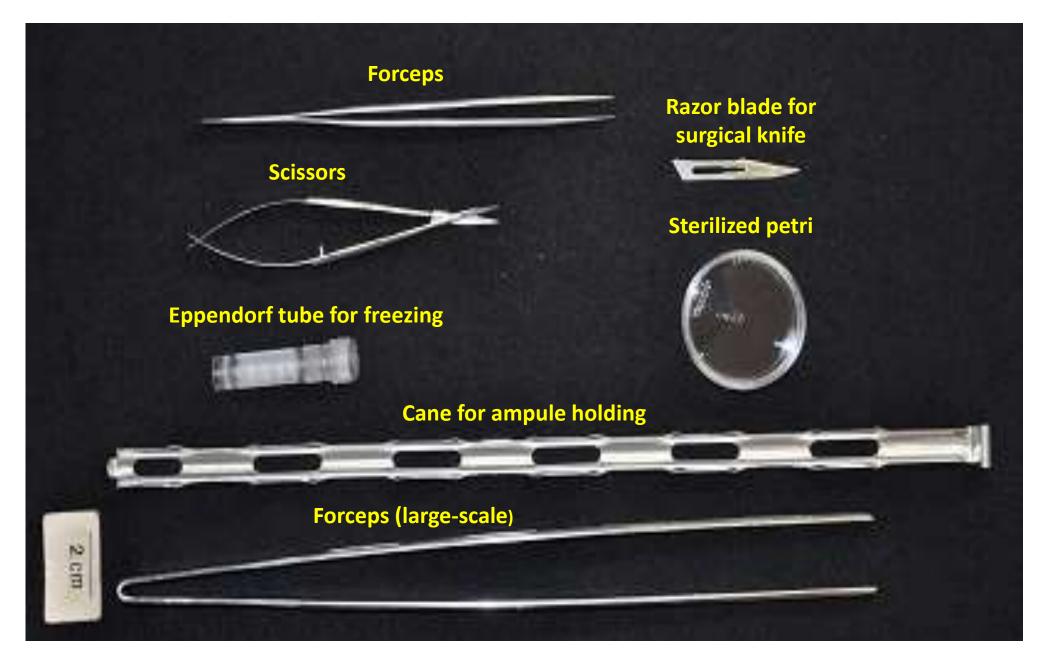
Manual for the cryopreservation of the silkworm germ cells

Yuuji Mochida, Yoko Takemura, Hiroshi Shinbo, Yutaka Banno

(Published in Sanshi-Konchu Biotec, Vol 83.)



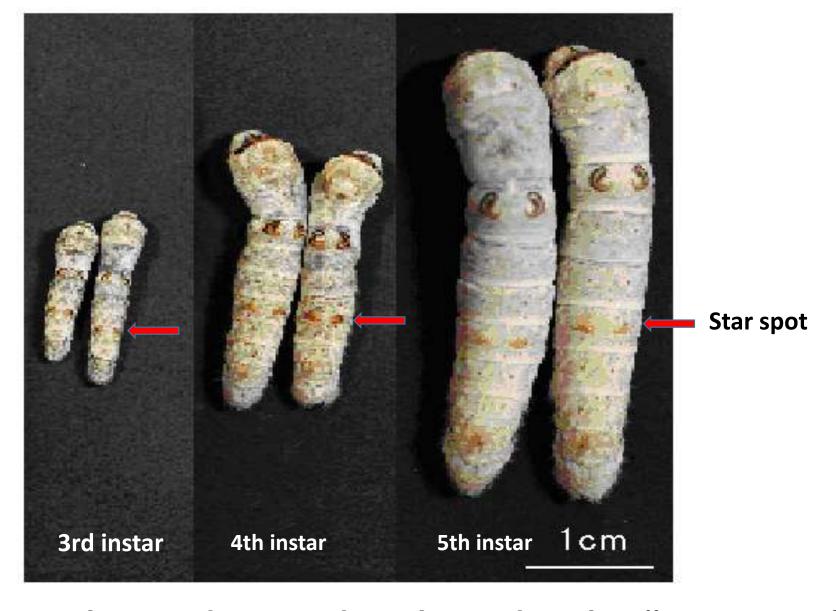
Procedure for cryopreserving the larval ovary



Lab wares used



Manipulation of collection and transplantation of ovaries should be done in clean benches



Ovaries are located immediately under the "star spots" on the 8th laraval segment



Sterilize body surface in the part of "star spots" with alcohol



Make small incisions on a pair of "star spots" with a razor blade

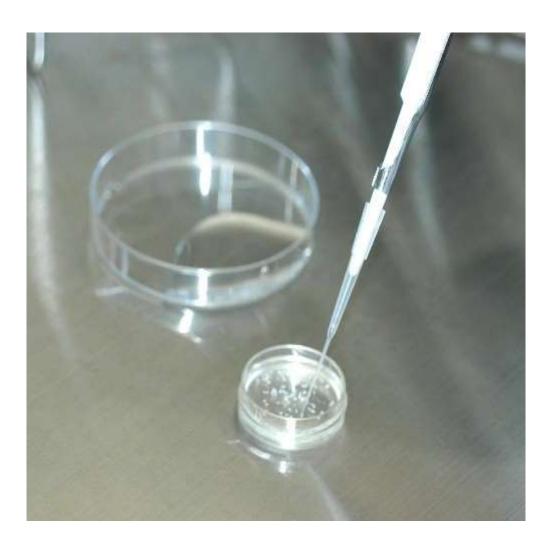


Extirpate ovaries on each side with forceps

Extirpate the larval ovaries



Collect the ovaries in Grace insect medium



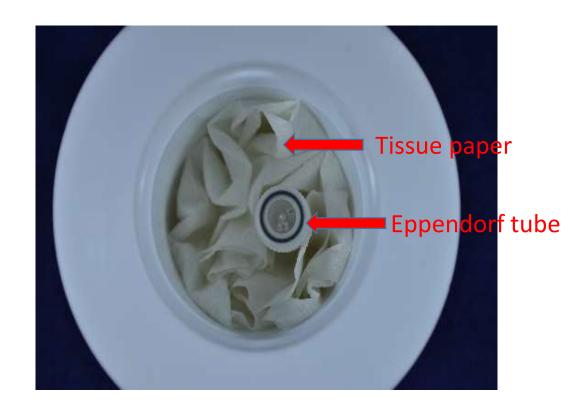
Suspend the collected ovaries in Grace insect medium containing 1.5M DMSO as cryoprotectant





Transfer the ovaries into Eppendorf tube for freezing by using forceps (A) or pipette (B)





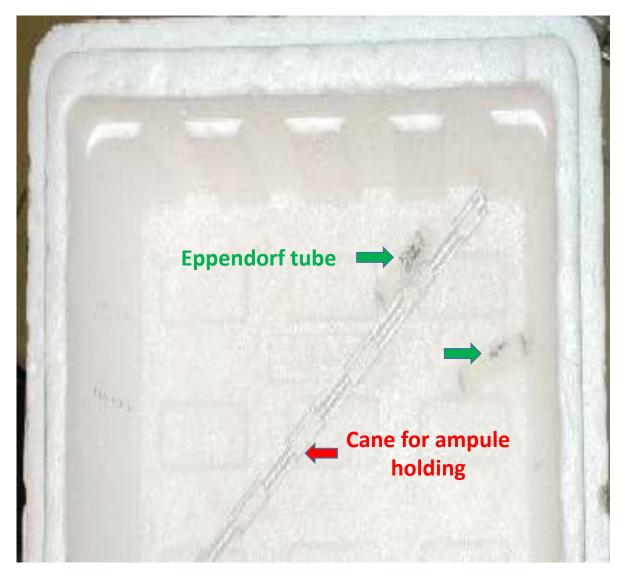
Fix the Eppendorf tube in BICELL (Bio freezing vessel)



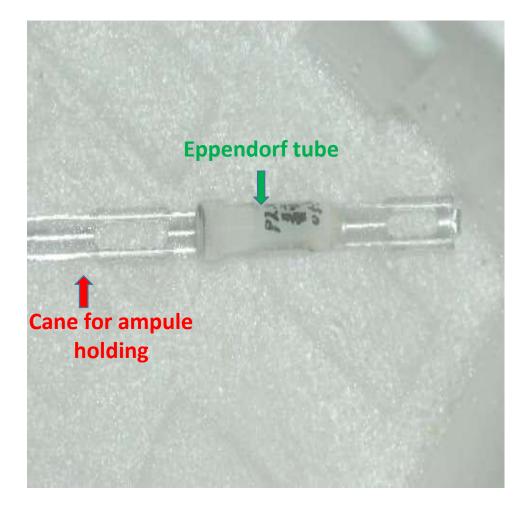


Freezer inside

Keep the BICELL (Bio freezing vessel) in -80°C freezer for ca. 3-20hrs



Fill box of Styrofoam with liquid nitrogen, and place cane for ampule holding in the liquid nitrogen.

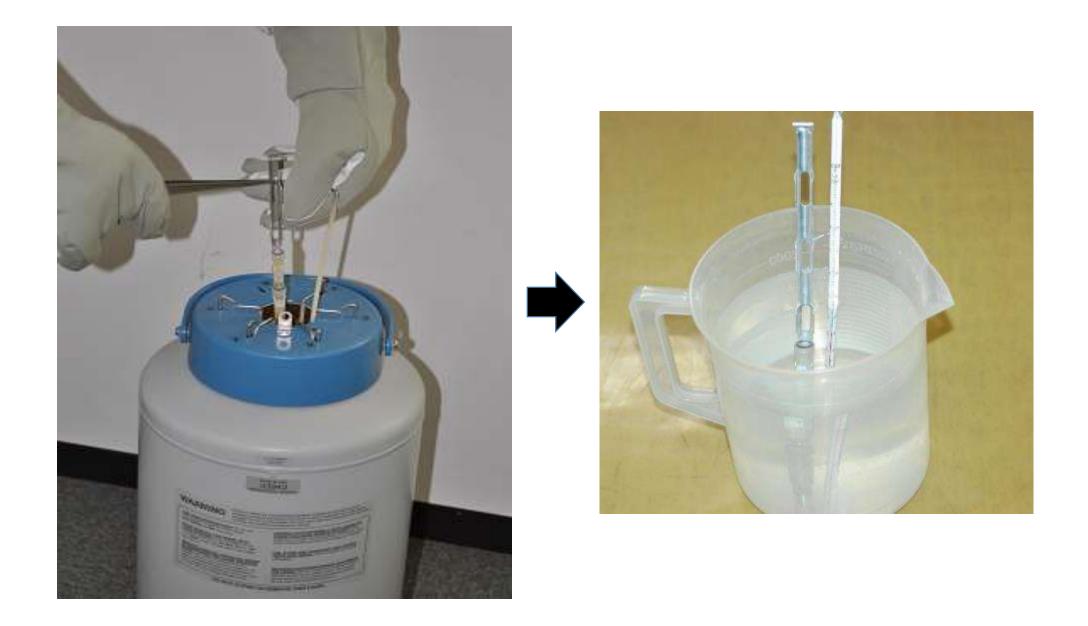


Then, attach the Eppendorf tube to the cane for ampule holding in liquid nitrogen by using forceps (large-scale)

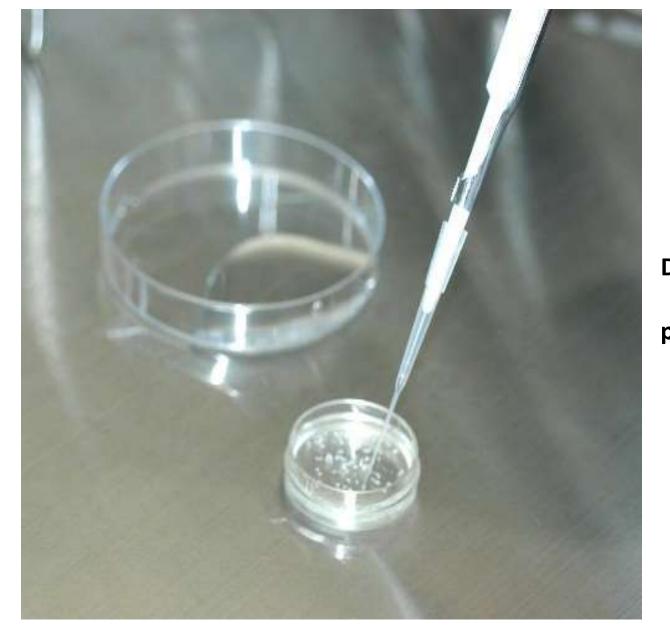


Mark the tip of the cane to distinguish samples

Store the cane in liquid nitrogen preserving container

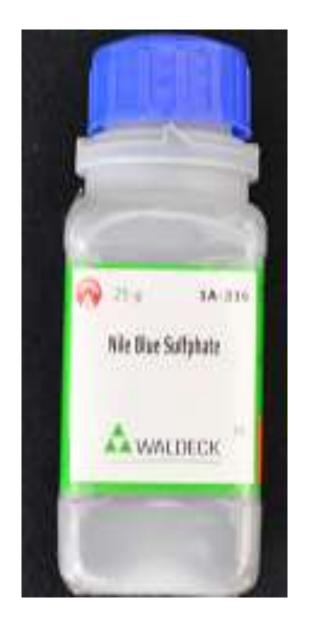


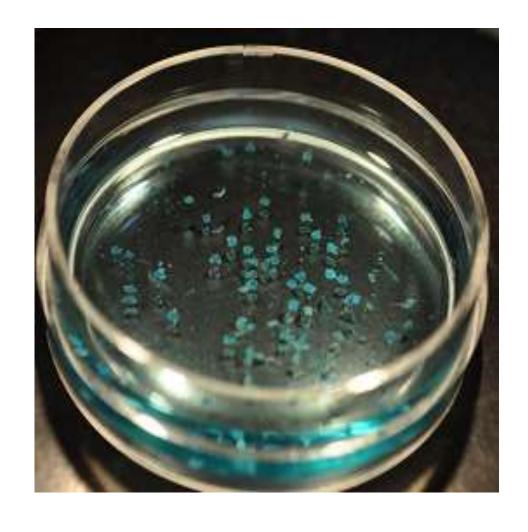
Sift the cane into a 37°C water bath to thaw the ovaries



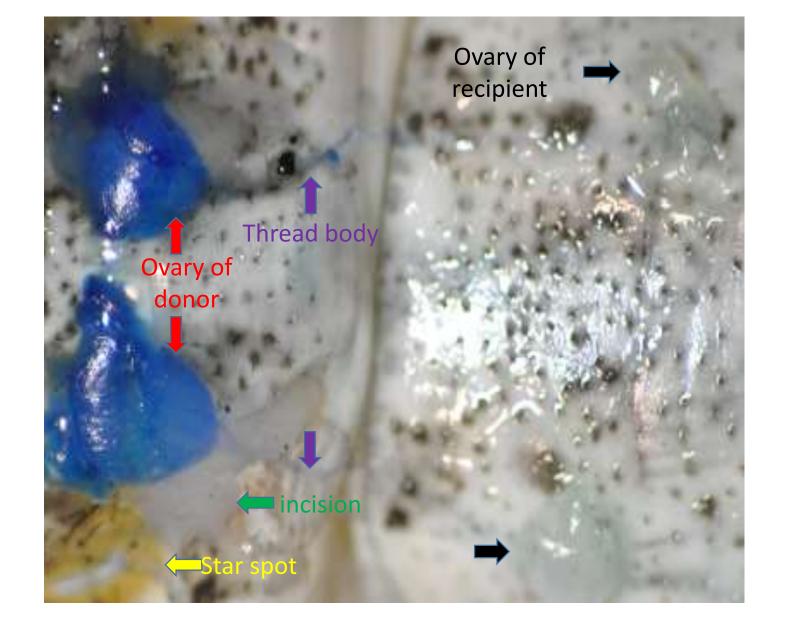
1.5M-DMSO into petri (upper). Then, pour Grace insect medium into the petri containing ovaries.

Replace the medium to remove DMSO (cryoprotectanat)

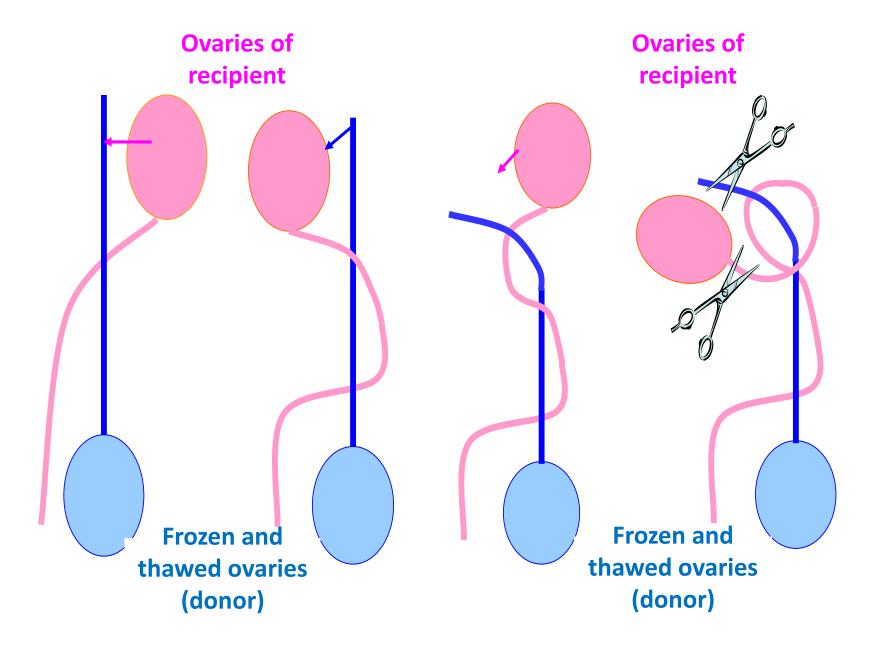




Stain the frozen and thawed ovaries with Nile Blue Sulphate



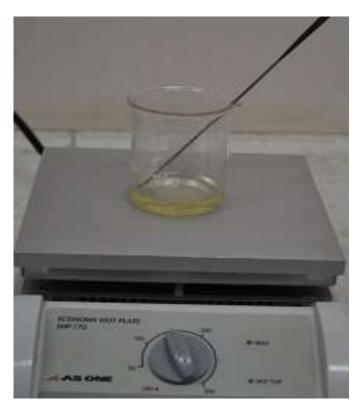
Ovaries of donor (blue) and recipient (milky white translucent)



Connect thread bodies of donor and those of recipient by coiling



Restore the donor ovary with the two thread bodies coiled around one another into the recipient via the original incision







Cover the incisions with paraffin





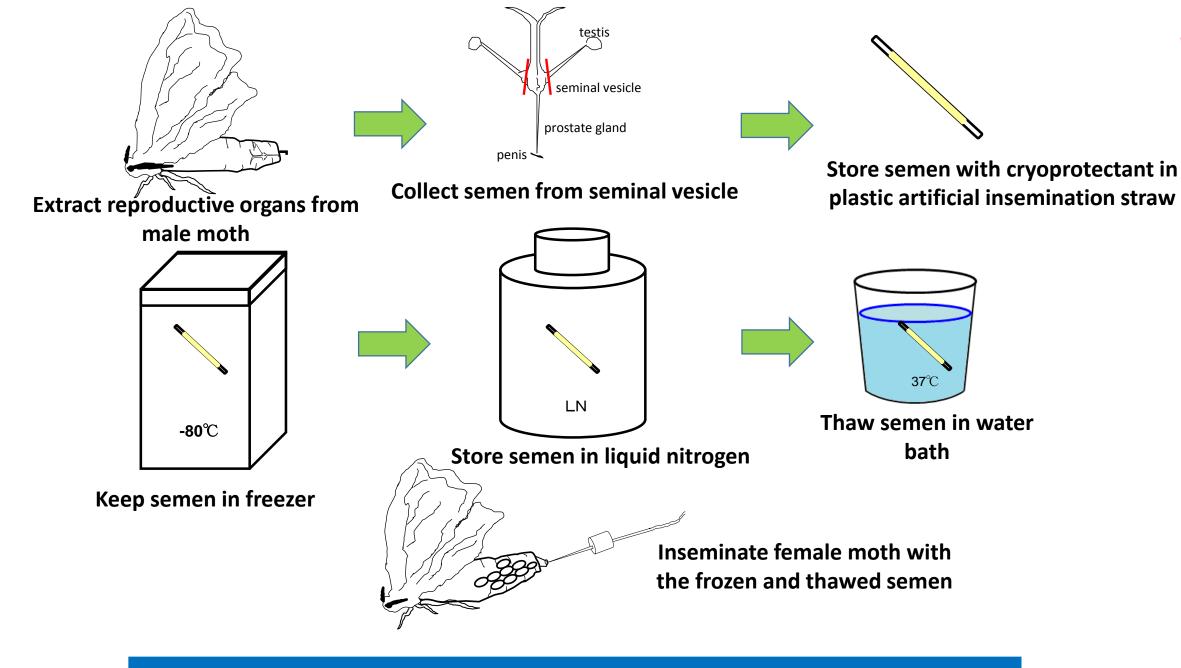


2 days after the transplantation

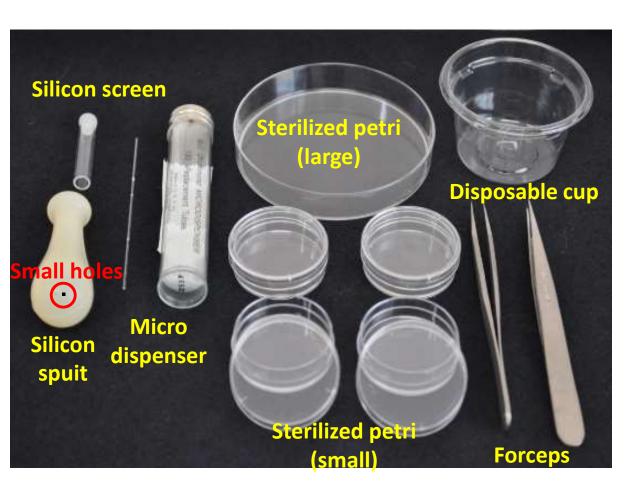
newly moulted 5th instar larvae

pupae

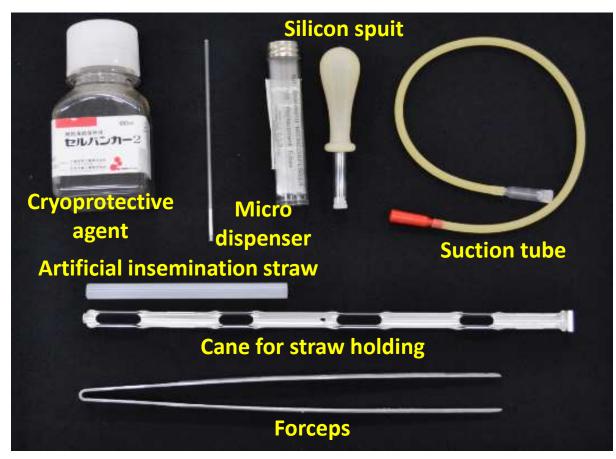
Rear the larvae to complete their life cycle



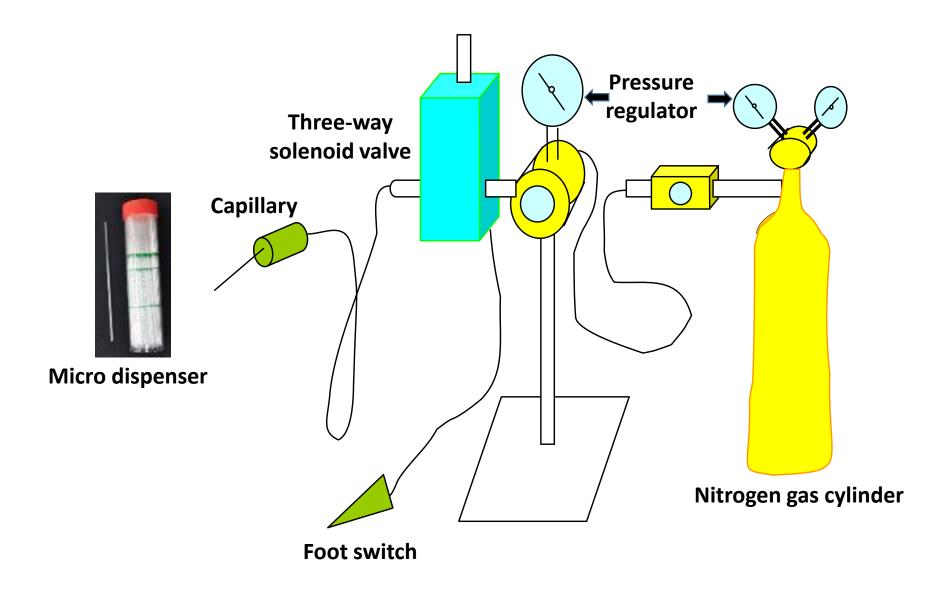
Procedure for cryopreserving the spermatozoa



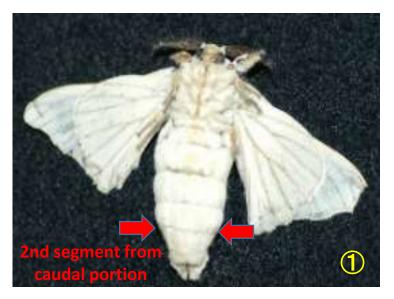




Lab wares used for cryopreseving semen



Instrument for artificial insemination





3

Anchor the ventral part of the 2nd segment with forceps

Tear up the segment

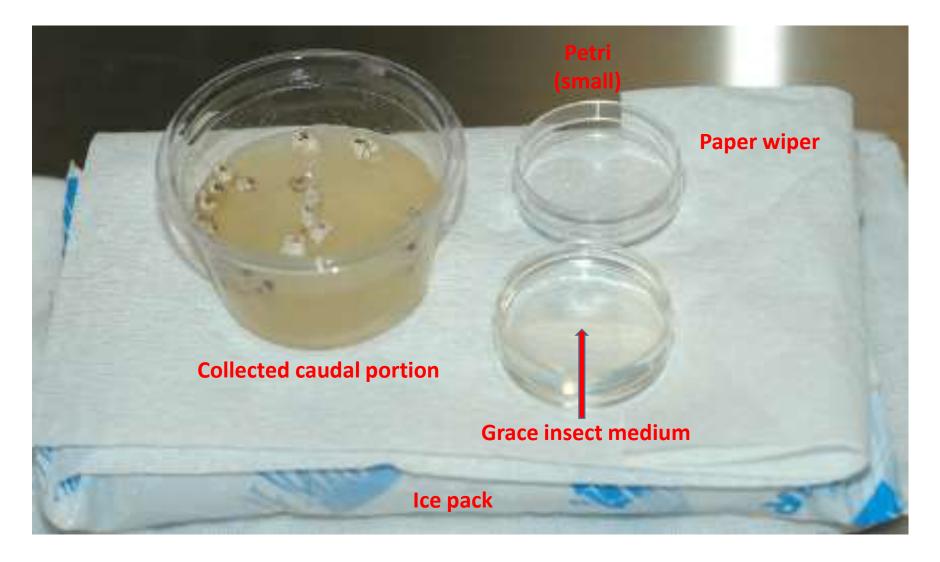


Internal whole reproductive organs containing seminal vesicle



Caudal portion containing whole internal reproductive organs

Collect caudal portion containing whole internal reproductive organs





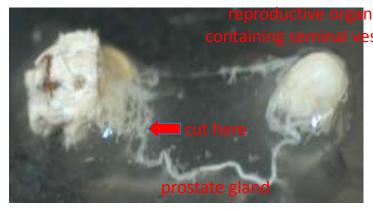
Prepare the collecting reproductive organs containing seminal vesicle



Fasten the innermost of the caudal portion softly with forceps



Hold the caudal portion down with the left forceps, and pull the reproductive organs softly with the right forceps

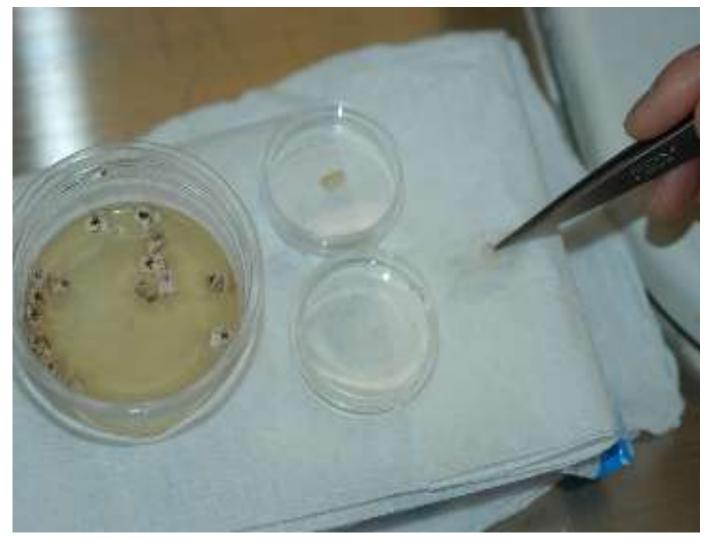






Cut the reproductive organs off at the tip of prostate gland

Collect the reproductive organs



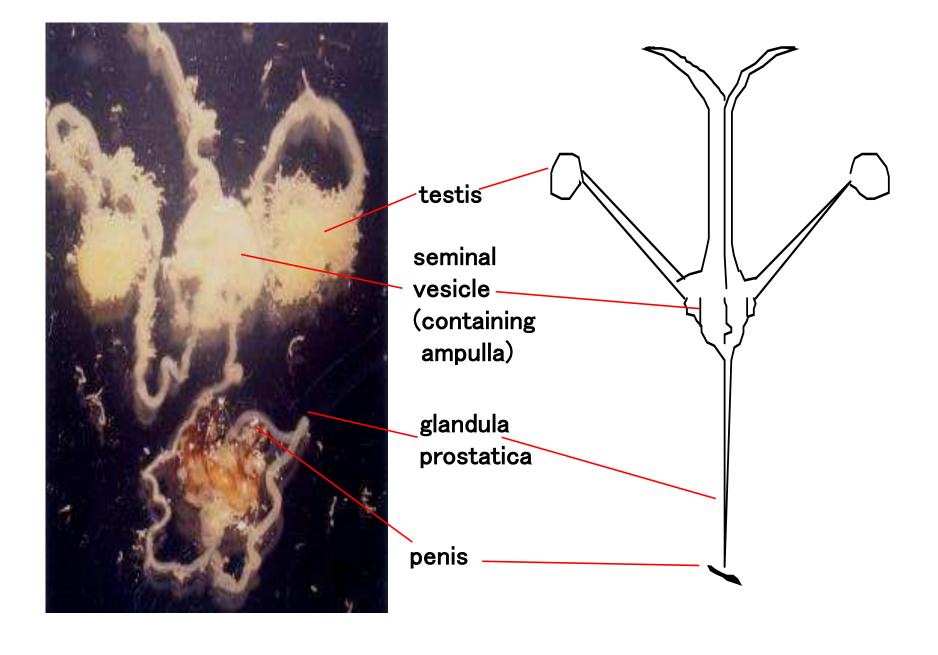


Wash the dirty things, like scaly hair, attached to the reproductive organs out in the petri filled with Grace insect medium

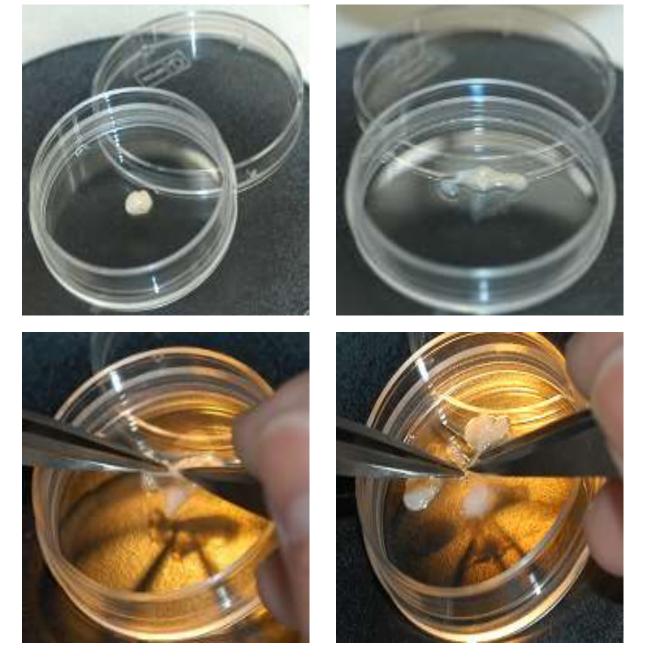
Wash the collected reproductive organs



Prepare the collecting semen



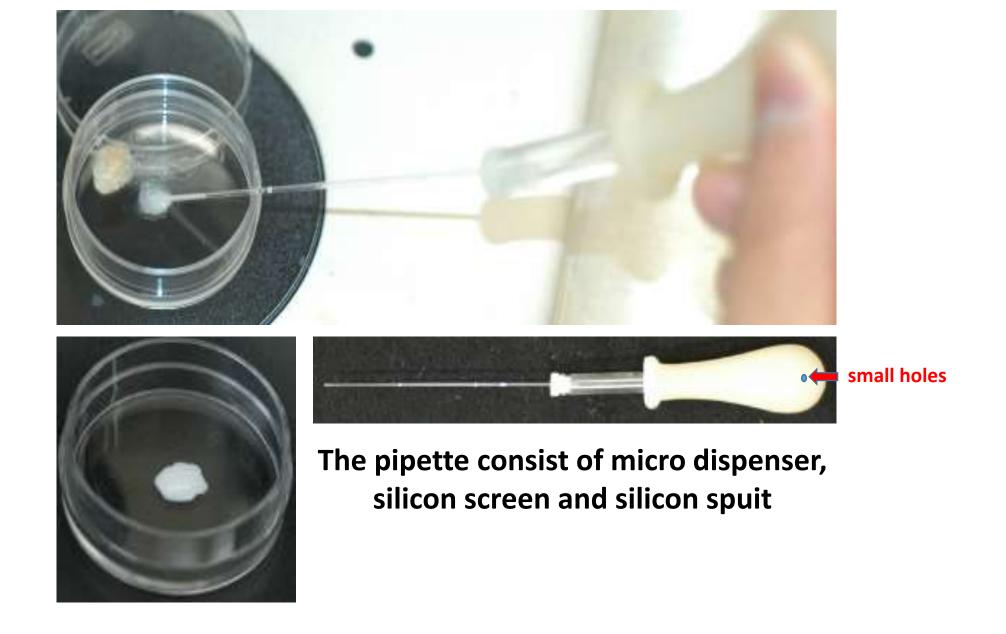
Reproductive organs of male moth



Spread the reproductive organs, and collect the semen by cutting the ampulla of the seminal vesicle off with forceps.

Cut the glandula prostatica off in the same manner to collect expressed prostatic secretion.

Collect the semen and expressed prostatic secretion

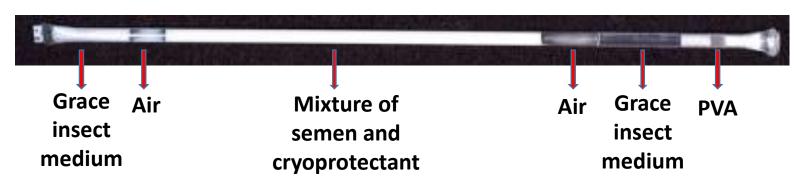


Collect the semen by using pipette



Store the semen in plastic artificial insemination straw by using suction tube





Store the semen in artificial insemination straw



Seal the both tips of the straw up with heat sealer







Freezer inside

Keep the straw in -80°C freezer for 10 min



Fill box of Styrofoam with liquid nitrogen. Then, place the straw frozen at -80°C in the liquid nitrogen



Then, attach the straw to the cane for straw holding in liquid nitrogen by using forceps (large-scale)



Store the cane in liquid nitrogen preserving container

Store the semen in liquid nitrogen



Shift the straw into 37°C water bath to thaw the semen



Cut the straw at the portion containing air, and collect the semen in petri

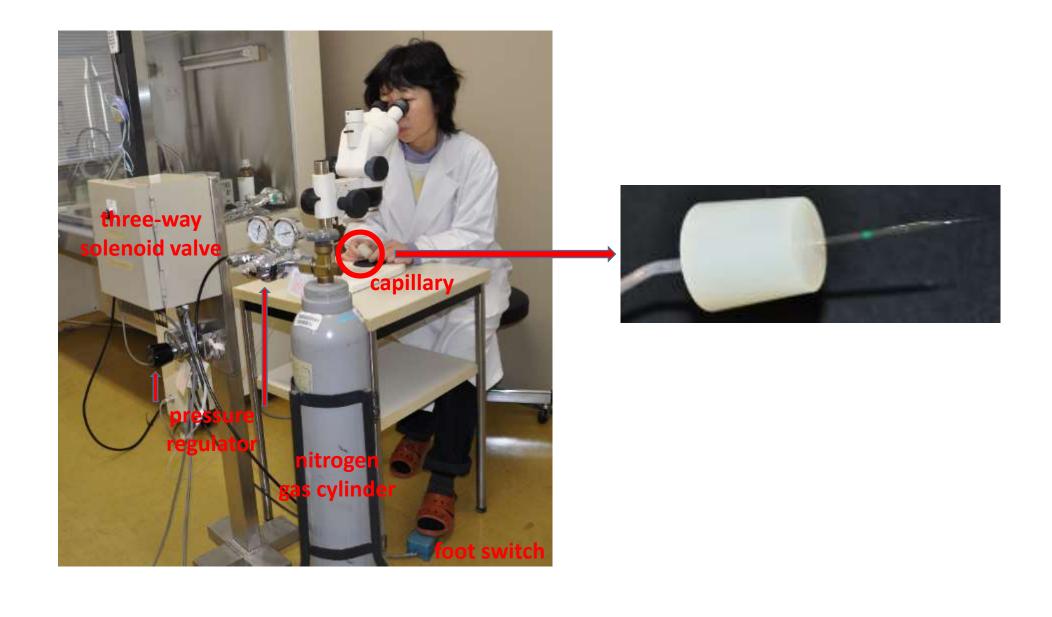
Thaw the semen stored in liquid nitrogen



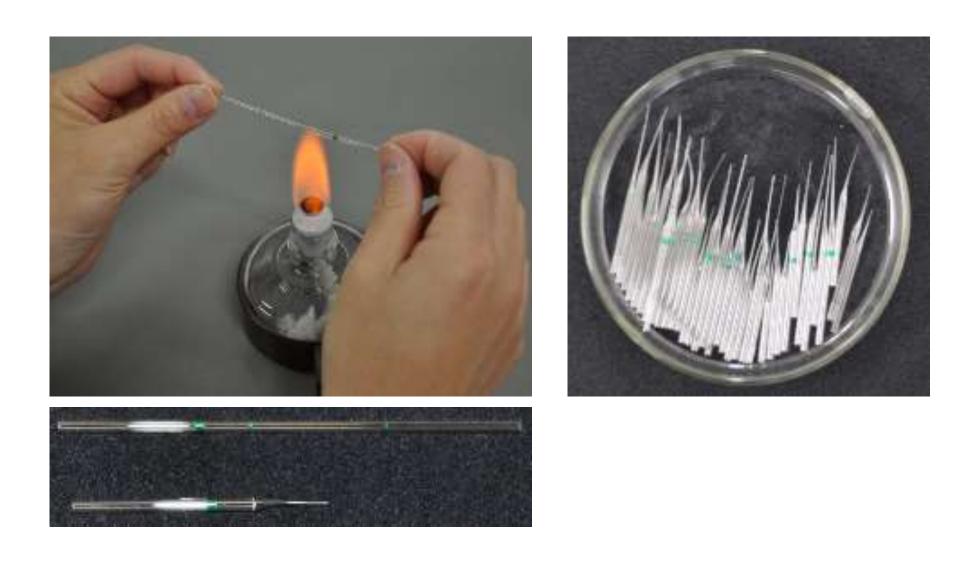




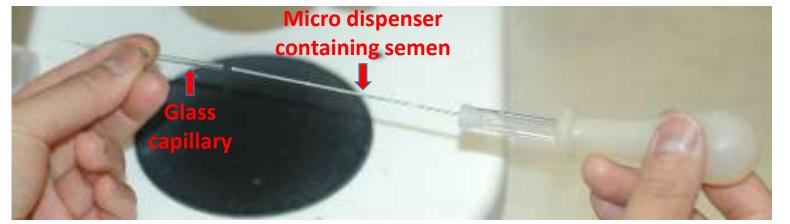
Mix the sperm with the trypsin solution at the time of artificial insemination

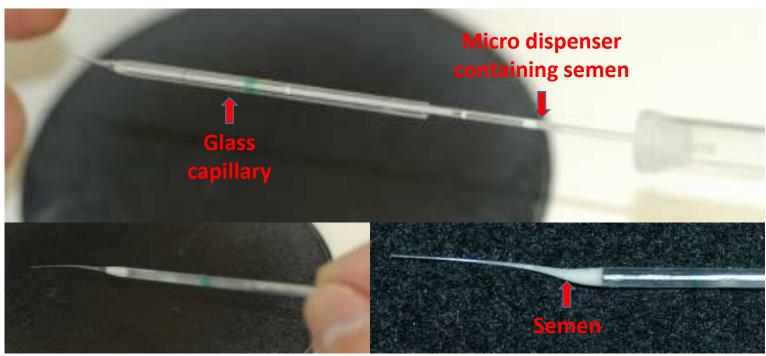


Artificial insemination work

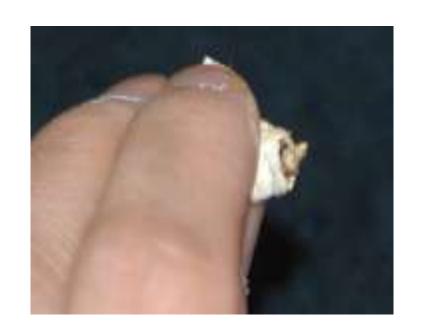


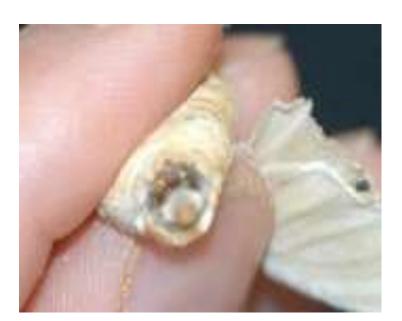
Make the glass capillary





Load the semen into glass capillary



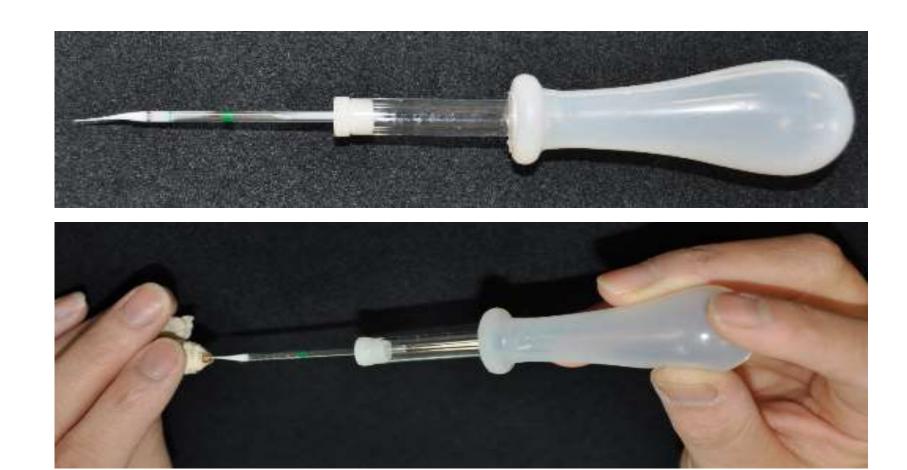




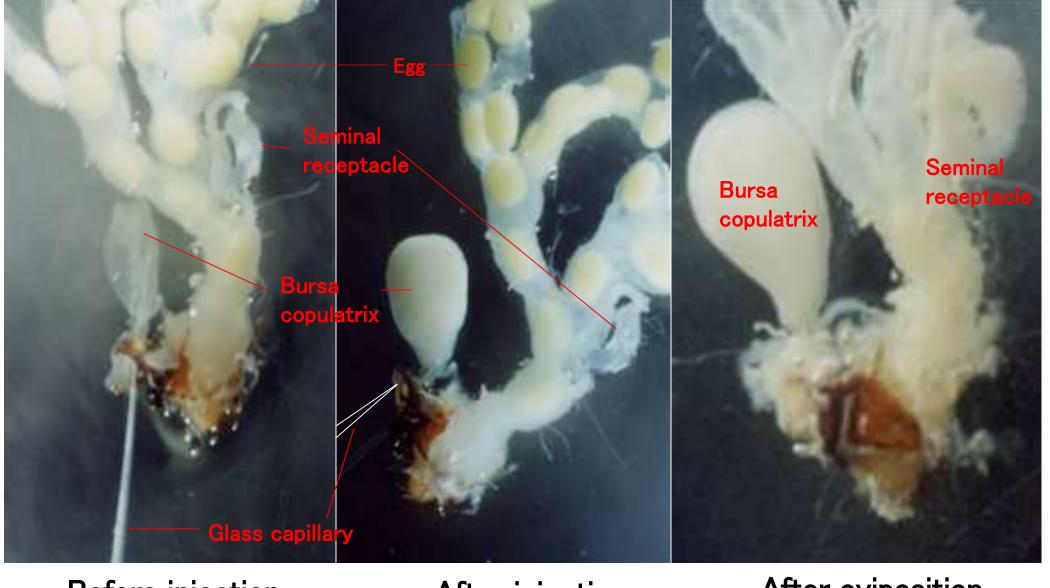


Inject an aliquot of semen into the bursa copulatrix of virgin female

Inject the semen into the bursa copulatrix



Manual operation for artificial insemination by using pipette

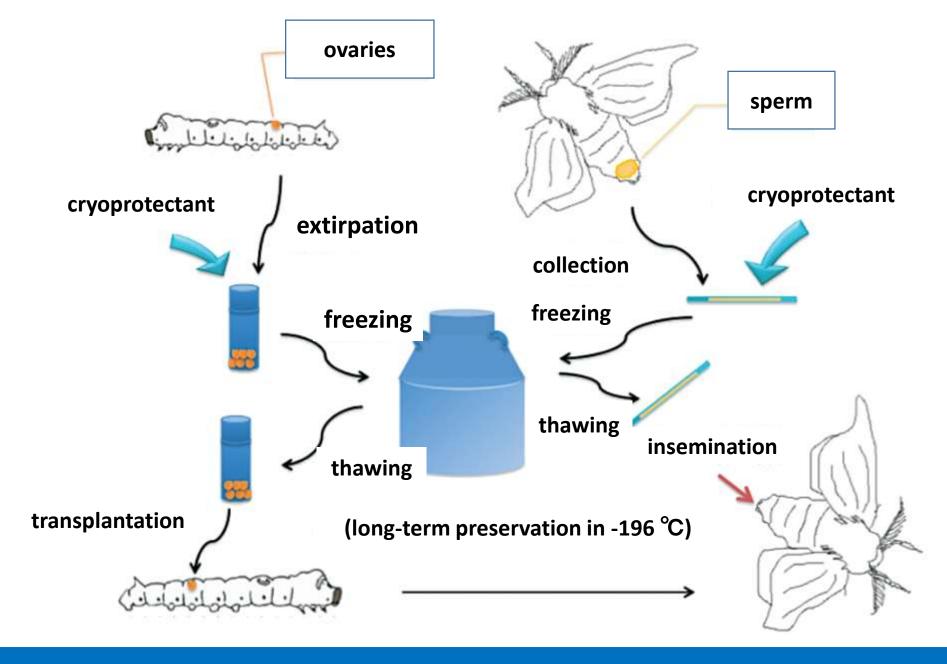


Before injection

After injection

After oviposition

Injection of semen into the bursa copulatrix of virgin female moth



Method for obtaining offspring using cryopreserved ovaries and sperm